

Template for Reporting Results of Biomarker Testing of Specimens From Patients With Diffuse Large B-Cell Lymphoma, Not Otherwise Specified (NOS)

Template web posting date: December 2014

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For the Members of the Cancer Biomarker Reporting Committee, College of American Pathologists

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CAP Diffuse Large B-Cell Lymphoma Biomarker Template Revision History

Version Code

The definition of the version code can be found at www.cap.org/cancerprotocols.

Version: DLBCL_Biomarkers 1.0.0.0

Summary of Changes

This is a new template.

Diffuse Large B-Cell Lymphoma Biomarker Reporting Template

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Completion of the template is the responsibility of the laboratory performing the biomarker testing and/or providing the interpretation. When both testing and interpretation are performed elsewhere (eg, a reference laboratory), synoptic reporting of the results by the laboratory submitting the tissue for testing is also encouraged to ensure that all information is included in the patient's medical record and thus readily available to the treating clinical team.

DIFFUSE LARGE B-CELL LYMPHOMA (DLBCL), NOT OTHERWISE SPECIFIED (NOS)

Select a single response unless otherwise indicated.

Note: Use of this template is optional.

+ SPECIMEN TYPE

- + ____ Peripheral blood
- + ____ Bone marrow
- + ____ Lymph Node (specify site): _____
- + ___ Other (specify): _____

RESULTS

- + Protein Expression (by immunohistochemistry [IHC] or flow cytometry) (Notes A and B)
- + ____ BCL2
 - + ___ Not detected
- + ____ CD5
 - + ____ Not detected
- + ____ CD20
 - + ___ Not detected
 - + ___ Detected
- + ____ CD30
 - + ___ Not detected
- + ____ Ki-67
 - + ____ Not detected
- + ____ MYC
 - + ___ Not detected
 - + ____ Detected
- + ___ Other marker(s) tested (specify): _____
 - + ___ Not detected
 - + ___ Detected

+ Subtype Classification (Note C)

- + ____ Germinal center-like
- + ____ Non-germinal center-like

+ Chromosomal Abnormalities (by fluorescence in situ hybridization [FISH]) (Note B)

- + ____ MYC rearrangement
 - + ____ Not detected
 - + ____ Detected
 - + ____ Other (specify): ______
- + ____ BCL2 rearrangement
 - + ____ Not detected
 - + ____ Detected
 - + ____ Other (specify): ______
- + ____ BCL6 rearrangement
 - + ____ Not detected
 - + ___ Detected
 - + ____ Other (specify): ___
- + Other probes tested (if applicable)
 - + Specify probe: _____
 - + Specify results: _____

+ Cytogenetic testing complete karyotype (specify): _____

+ Somatic Gene Mutations (by sequencing) (Note D)

- + ____ Not detected
- + ___ Detected (specify variant): _____
- + ____ Other (specify): _____

+ Other Markers Tested (if applicable)

- + Specify marker: _____
- + Specify results: _____

+ METHODS

+ Protein Expression (select if tested) (Notes A and B)

- + ____ IHC
 - + ____ BCL2 (specify clone): _____
 - + ___ CD5 (specify clone): _____
 - + ___ CD20 (specify clone): _____
 - + ___ CD30 (specify clone): _____
 - + ____ Ki-67 (specify clone): ______
 - + ____ MYC (specify clone): ______
 - + ___ Other(s) (specify clone): _____
- + ____ Flow cytometry
 - + ____ BCL2 (specify clone): _____
 - + ____ CD5 (specify clone): _____
 - + ___ CD20 (specify clone): _____
 - + ___ CD30 (specify clone): _____
 - + ___ Other(s) (specify clone): _____

- + ____ FISH
 - + BCL2 probe
 - + ____ Break apart
 - + ____ Fusion
 - + BCL6 probe
 - + ____ Break apart
 - + ____ Fusion
 - + MYC probe
 - + ____ Break apart
 - + ____ Fusion

+ Subtype Classification (Note C)

- + ____ Hans (CD10, BCL-6, MUM1)
- + ___ Choi (GCET1, CD10, MUM1, BCL6, FOXP1)
- + ____ Tally (CD10, GCET1, MUM1, FOXP1, LMO2)
- + ___ Gene expression profiling (specify platform/method): _____
- + ___ Other (specify): _____

+ Gene Sequencing (Note D)

- + Gene sequencing platform (specify):
- + Maximum sensitivity: ______ (variant allele frequency) + Genes/exons sequenced (specify): ______

Explanatory Notes

A. Protein Expression

The antibodies listed are included in the template because they have therapeutic or prognostic significance.

CD20 assessment is mandatory for therapeutic planning because the standard therapy for diffuse large B-cell lymphoma (DLBCL) patients is R-CHOP (rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone).¹ Knowledge of CD20 expression is therefore recommended as a justification for using rituximab.

CD30 assessment is recommended because of the potential utility of the anti-CD30 antibody drug conjugate, brentuximab vedotin. Approximately 10% to 15% of DLBCL cases express CD30, and these patients may be eligible for this agent if they fail standard therapy.²

CD5 assessment is thought to be of prognostic value because a small subset (5%-10%) of DLBCL cases. Patients with CD5+ DLBCL have a more aggressive clinical course. These patients tend to be older and have elevated serum LDH levels, poorer performance status, and a higher frequency of central nervous system involvement.³

MYC assessment by immunohistochemistry (IHC) is of prognostic value and needs to be evaluated in conjunction with conventional cytogenetic analysis or FISH to assess chromosome locus 8q24/MYC rearrangements. Approximately 10% of DLBCL cases carry MYC translocations, and up to 30% to 40% of cases overexpress MYC by IHC, with positivity defined in various studies as >40% or >50% positive cells. Therefore, MYC can be overexpressed via mechanisms other than translocation.^{4,5}

The combination of MYC and BCL2 and/or BCL6 gene rearrangements as shown by conventional cytogenetic or FISH analysis is known as double (or triple) hit lymphoma. Patients with this combination of abnormalities have a poor prognosis.⁶

MYC positivity by IHC may be useful as a screen for MYC translocations as it is rare for a translocation positive case to be negative for MYC by IHC. MYC expression combined with BCL2 overexpression is also associated with a poorer prognosis (so-called IHC double hit lymphoma).

BCL2 assessment also has prognostic value and needs to be evaluated in conjunction with conventional cytogenetic analysis or FISH to assess chromosome locus 8q24/MYC rearrangements. In patients treated with CHOP, BCL2 overexpression correlates with poorer prognosis in the germinal center type of DLBCL.⁷ BCL2 overexpression combined with MYC overexpression correlates with a poorer prognosis (IHC double hit lymphoma).

B. Fluorescence In Situ Hybridization (FISH)

Recent studies have demonstrated that DLBCL with rearrangements of MYC and BCL2 or BCL6 comprise a distinct subgroup of cases, often termed double hit lymphomas, characterized by overlapping morphologic features with Burkitt lymphoma and a more aggressive clinical course.⁸⁻¹¹

C. Subtyping

Studies have shown there are prognostic differences in DLBCL that are germinal center derived (GC) versus non-germinal center derived (NGC). Several methodologies have been proposed for predicting GC versus NGC derivation.¹² In general, DLBCL of GC type is associated with a better prognosis. The most commonly applied immunohistochemical methodologies, which serve as a substitute for gene expression arrays (a gold standard for GC versus NGC), are Hans classifier,¹³ Choi classifier,¹⁴ and Tally classifier.¹⁵

D. Sequencing

Somatic variants in TP53, MYD88, PAX5, TNFRSF14, and other genes have been shown to correlate with cell of origin, patient outcome, and diagnosis in some studies. When such variants are identified they should be reported according to the Human Genome Variant Society (HCVS) guidelines nomenclature.

References

- 1. Coiffier B. Treatment of diffuse large B-cell lymphoma. Curr Hematol Rep. 2005;4(1):7-14.
- 2. Hu S, Xu-Monette ZY, Balasubramanyam A, et al. CD30 expression defines a novel subgroup of diffuse large B-cell lymphoma with favorable prognosis and distinct gene expression signature: a report from the International DLBCL Rituximab-CHOP Consortium Program Study. *Blood*. 2013;121(14):2715-2724.
- 3. Jain P, Fayad LE, Rosenwald A, et al. Recent advances in *de novo* CD5+ diffuse large B cell lymphoma. *Am J Hematol.* 2013;88(9):798-802.
- 4. Johnson NA, Slack GW, Savage KJ, et al. Concurrent expression of MYC and BCL2 in diffuse large Bcell lymphoma treated with rituximab plus cyclophosphamide, doxorubicin, vincristine, and prednisone. J Clin Oncol. 2012;30(28):3452-3459.
- 5. Hu S, Xu-Monette ZY, Tzankov A, et al. MYC/BCL2 protein coexpression contributes to the inferior survival of activated B-cell subtype of diffuse large B-cell lymphoma and demonstrates high-risk gene expression signatures: a report from The International DLBCL Rituximab-CHOP Consortium Program. *Blood*. 2013;121(20):4021-4431.
- 6. Aukema SM, Siebert R, Schuuring E, et al. Double-hit B-cell lymphomas. Blood. 2011;117(8):2319-2331.
- Iqbal J, Meyer PN, Smith LM, et al. BCL2 predicts survival in germinal center B-cell-like diffuse large Bcell lymphoma treated with CHOP-like therapy and rituximab. *Clin Cancer Res.* 2011 Dec 15;17(24):7785-7795.
- 8. Niitsu N, Okamoto M, Miura I, Hirano M. Clinical features and prognosis of de novo diffuse large Bcell lymphoma with t(14;18) and 8q24/c-MYC translocations. *Leukemia*. 2009;23(4):777-783.
- 9. Aukema SM, Siebert R, Schuuring E, et al. Double-hit B-cell lymphomas. Blood. 2011;117(8):2319-2331.
- Li S, Lin P, Fayad LE, et al. B-cell lymphomas with MYC/8q24 rearrangements and IGH@BCL2/t(14;18)(q32;q21): an aggressive disease with heterogeneous histology, germinal center B-cell immunophenotype and poor outcome. Mod Pathol. 2012;25(1):145-156.
- 11. Lin P, Medeiros LJ. The impact of MYC rearrangements and "double hit" abnormalities in diffuse large B-cell lymphoma. *Curr Hematol Malig Rep.* 2013;8(3):243-252.
- 12. Alizadeh AA, Eisen MB, Davis RE, et al. Distinct types of diffuse large B-cell identified by gene expression profiling. *Nature*. 2000;403(6769):503-511.
- Hans CP, Weisenburger DD, Greiner TC, et al. Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a tissue microarray. Blood. 2004;103(1):275-282.
- 14. Choi WW, Weisenburger DD, Greiner TC, et al. A new immunostain algorithm classifies diffuse large B-cell lymphoma into molecular subtypes with high accuracy. *Clin Cancer Res*. 2009;15(17):5494-5502.
- Meyer PN, Fu K, Greiner TC, Smith LM, et al. Immunohistochemical methods for predicting cell of origin and survival in patients with diffuse large B-cell lymphoma treated with rituximab. J Clin Oncol. 2011;29(2):200-207.